Acquired Amegakaryocytic Thrombocytopenic Purpura Progressing into Aplastic Anemia

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Received August 10, 2017; Accepted December 5, 2017.

Key words: Acquired amegakaryocytic thrombocytopenic purpura – Pure megakaryocytic aplasia – TPO agonist – Aplastic anemia

Abstract: Acquired amegakaryocytic thrombocytopenic purpura (AATP) is a rare hematological disorder characterized by severe thrombocytopenia and a complete or near-to complete absence of megakaryocytes in the bone marrow, while granulopoiesis, as well as erythropoiesis are usually preserved. Although autoimmune mechanisms are believed to be causative, the exact underlying pathogenesis is not known. To date, only few cases have been reported and management of this disease remains controversial with immunosuppression being the treatment modality of choice in the majority of patients. In this article, we report a case of newly acquired AATP without an associated autoimmune disease, refractory to corticoids, intravenous immunoglobulin (IVIG) and second-generation TPO (thrombopoietin) agonists, which have recently been approved for the treatment of thrombocytopenia. Finally, in accordance with other reports, disease progression into aplastic anemia has occurred.

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https://doi.org/10.14712/23362936.2017.16
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Introduction
Acquired amegakaryocytic thrombocytopenic purpura (AATP), also known as acquired pure megakaryocytic aplasia, is a rare hematological disorder reported in patients ranging from 2 to 89 years of age with predilection for males in the age group above 60 years (Figure 1). It can either occur isolated or be associated with systemic lupus erythematosus (SLE) (Cela et al., 2010; Ernestho-ghoud et al., 2015) and other autoimmune diseases (Hashimoto et al., 2016), as well as hematological malignancies such as non-Hodgkin lymphoma (Lugassy, 1996).

Most patients with this disease are initially diagnosed with immune thrombocytopenic purpura (ITP) and receive treatment with corticosteroids (either prednisone or dexamethasone) until a bone marrow aspirate or biopsy is performed, which reveals normal erythro- and granulopoiesis but near to complete absence of megakaryocytes (Hoffman et al., 1982), usually. Dysplastic changes consistent with myelodysplastic syndromes are not seen initially, but may appear in the clinical course of the disease with progression to myelodysplastic syndrome (Erkurt et al., 2005). Furthermore, progression to aplastic anemia (Meytes et al., 1984; King et al., 1997) and association with hematologic malignancies (Geissler et al., 1987), as well as presence of concurrent anemia has been reported (Niparuck et al., 2008).

The exact underlying pathogenic mechanisms remain unknown to some extent, but successful treatment with immunosuppressive agents points towards an autoimmune-mediated process. The variable outcome with different treatments, however, is indicative of not only a singular mechanism of pathogenesis. Indeed, inhibition at the level of the thrombopoietin receptor (Hoffman et al., 1989) due to a humoral agent, antibodies against thrombopoietin itself (Katsumata et al., 2003) or a T-cell mediated process (Gewirtz et al., 1986; Doubek et al., 2006) have been described.

Figure 1 – Graphical representation of age and gender distribution of 30 patients with diagnosed AATP.
In contrast to that, congenital amegakaryocytic thrombocytopenia (CAMT) is characterized by a mutation in the \textit{MPL} gene coding for the TPO (thrombopoietin) receptor and is inherited in an autosomal recessive manner. Depending on the type of mutation, CAMT can be further classified into type-I or type-II according to the type of underlying mutation predicted to result in either complete loss of function or some retained function of the TPO receptor (nonsense and missense mutation, respectively) (King et al., 2005). However, CAMT without mutation in the \textit{MPL} gene has been described in association with other diseases (e.g. Hoyeraal-Hreidarsson syndrome) and is referred to as type-III (King et al., 2004).

To date, allogeneic hematopoietic stem cell transplantation remains the only therapeutic option, whilst gene therapy or TPO agonists binding to partially functioning TPO receptors might provide benefit in the future.

Finally, AATP and CAMT have to be, amongst other things, distinguished from ITP, which is characterized by either an increased degradation of thrombocytes due to autoantibodies (Cines et al., 2009), inhibition of thrombopoiesis by autoantibodies interfering with megakaryopoiesis (Chang et al., 2003; McMillan et al., 2004), T-cell mediated toxicity towards thrombocytes (Olsson et al., 2003) or relatively insufficient thrombopoietin concentration (Emmons et al., 1996).

No primary therapy for AATP has been established to date. Immunosuppression, however, remains the mainstay of therapy. Treatments shown to be successful include the use of corticoids (Sakurai et al., 1984), intravenous immunoglobulins (IVIG) (Leach et al., 1999), cyclosporine (Omri et al., 2010), anti-thymocyte globulin (ATG) (Niparuck et al., 2008), as well as allogeneic stem cell transplantation (Lonial et al., 1999) and anti-CD20 antibodies (Deeren and Dorpe, 2010; Mirzania et al., 2014). While these and other therapeutic agents have shown a wide range of response rates, reports emerged reporting success with thrombopoietin (TPO) receptor agonists recently; a treatment option that has already been theorized earlier (Lown et al., 2010). Published case reports include the use of Elthrombopaoq in a patient with AATP and associated systemic lupus erythematosus (SLE) (Cela et al., 2010), as well as Romiplostim in a patient without underlying autoimmune disease (Shigekiy et al., 2015).

\textbf{Material and Methods}

The literature review was performed with searches on google scholar using the keywords “acquired amegakryocytic”, “acquired pure megakaryocytic aplasia”. Statistical analysis was performed on 30 patients (including the patient from this report) published in 19 articles. For this, only publications and case reviews reporting adult patients without significant comorbidities and autoimmune disease have been included. Gender, age and treatment modality have been analysed using Excel. Patients receiving simultaneous treatment with more than one agent have been excluded from response rate analysis.
Case report
A 61-year-old Caucasian male with a history of recent epistaxis, easy bruising and petechiae, which developed suddenly two weeks before admission. The patient history includes a perforated sigma diverticulitis 9 years ago, a total endoprothesis of the knee 2006, as well as a peripheral arterial occlusive disease stage IIa, adiposity with a BMI (body mass index) of 35 and a nicotine abuse of 45 pack years. No allergies or intolerances are known. On admission in a peripheral hospital due to severe epistaxis, blood workup showed marked thrombocytopenia of 0/nl and makrocytic, hyperchromic anemia (10.2 g/dl; Table 1), which worsened progressively with values reaching 7.4 g/dl five days later, probably due to several episodes of intermittent epistaxis. EDTA (ethylendiaminetetraacetic acid) induced pseudothrombocytopenia was ruled out.

Serologically there was no evidence of a hepatitis B, C or HIV infection. Vitamin $B_{12}$ concentration was normal, folic acid was mildly reduced at 4 ng/l and substitution was started until concentration above the upper normal limit was detected. Autoimmune diagnostics showed normal lupus anticoagulans concentration as well as a normal lupus sensitive – apTT. No thrombocyte antibodies were detected, coombs test was negative. An ultrasound of the abdomen showed no splenomegaly and lactate dehydrogenase (LDH) was within normal limits.

Suspecting a case of immune thrombocytopenia the patient initially received a prednisone burst with 250 mg/day and, due to refractory thrombocytopenia, subsequently 80 g intravenous immunoglobulins, which did not lead to a rise in

Table 1 – Initial and subsequent blood counts

<table>
<thead>
<tr>
<th></th>
<th>Initially</th>
<th>1 month later</th>
<th>Approx.</th>
<th>Approx.</th>
<th>Unit</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>14 months later</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hemoglobin</td>
<td>10.2</td>
<td>8.4</td>
<td>8.9*</td>
<td></td>
<td>g/dl</td>
</tr>
<tr>
<td>Hematokrit</td>
<td>30.0</td>
<td>0.24</td>
<td>0.25</td>
<td></td>
<td>l/l</td>
</tr>
<tr>
<td>Erythrocytes</td>
<td>3.1</td>
<td>2.5</td>
<td>3.1</td>
<td></td>
<td>/pl</td>
</tr>
<tr>
<td>MCH</td>
<td>33.0</td>
<td>34.0</td>
<td>29.0</td>
<td></td>
<td>pg</td>
</tr>
<tr>
<td>MCHC</td>
<td>34.0</td>
<td>35.0</td>
<td>36.0</td>
<td></td>
<td>g/dl</td>
</tr>
<tr>
<td>MCV</td>
<td>98.0</td>
<td>97.0</td>
<td>81.0</td>
<td></td>
<td>fl</td>
</tr>
<tr>
<td>Leukocytes</td>
<td>6.05</td>
<td>4.46</td>
<td>0.57</td>
<td></td>
<td>/nl</td>
</tr>
<tr>
<td>Thrombocytes</td>
<td>0.0</td>
<td>8.0</td>
<td>22.0*</td>
<td></td>
<td>/nl</td>
</tr>
<tr>
<td>Neutrophils</td>
<td>64.0</td>
<td>59.2</td>
<td></td>
<td></td>
<td>%</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>22.0</td>
<td>29.4</td>
<td></td>
<td></td>
<td>%</td>
</tr>
<tr>
<td>Monocytes</td>
<td>9.3</td>
<td>7.5</td>
<td></td>
<td></td>
<td>%</td>
</tr>
<tr>
<td>Eosinophils</td>
<td>4.5</td>
<td>1.3</td>
<td></td>
<td></td>
<td>%</td>
</tr>
<tr>
<td>Basophils</td>
<td>0.2</td>
<td>0.0</td>
<td></td>
<td></td>
<td>%</td>
</tr>
</tbody>
</table>

*transfusions received; MCH – mean corpuscular hemoglobin; MCHC – mean corpuscular hemoglobin concentration; MCV – mean corpuscular volume

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thrombocyte counts. Due to symptomatic anemia the patient received several erythrocyte transfusions. He was then referred to our clinics for further diagnostics and treatment.

Upon admission we decided to try another cycle of corticoids (Dexamethasone 40 mg/day, 4 days) and intravenous immunoglobulins at a concentration of 1 g/kg body weight (BW) (110 g) to confirm the refractoriness to these immunosuppressive agents. We also performed bone marrow cytology, flow cytometry of bone marrow blood and a bone marrow biopsy with subsequent histological examination, as well as chromosomal analysis. The cytology showed a near to complete absence of megakaryocytes with preserved granulo- and erythropoiesis. Slight dysplastic changes such as minimal misshaped nuclei and pseudo-pelger cells could be seen upon cytological examination. Ring sideroblasts were not observed.

Repeated chromosomal analysis showed normal male karyotype XY, 46. Flow-cytometry revealed no lead towards myelodysplasia and histological examination of the bone marrow biopsy confirmed the presence of normal maturation of the white and red lineage with nearly total absence of megakaryocytes as well as minimal increase in interstitial reticular fibers. There was no sign of an intramedullary increase in CD34+ cells and no infiltration by lymphoid cells. Erythropoietin concentration was well above normal limits.

During the in-hospital stay the patient developed fever with temperatures around 39 °C, which was initially treated with Tazobactam/Piperacillin i.v. and then escalated to Meropenem i.v., and empirical Caspofungin i.v., as well as Clarithromycin p.o. after several days in a stepwise manner prior to further diagnostics due to suspicion of atypical pneumonia. Blood cultures showed no bacterial growth. In order to localize the focus of the suspected infection a CT (computed tomography) scan of the chest was performed which showed no pneumonia, but incidentally revealed an 2×1.4 cm adrenal adenoma with partially negative Hounsfield units. The fever resolved during the course of treatment and

![Figure 2 – Thrombocyte and leukocyte counts during therapy.](image)
C-reactive protein concentration decreased accordingly so that antibiotic and antimycotic treatment was discontinued.

In summary of the results we then diagnosed the patient with acquired amegakaryocytic thrombocytopenic purpura. Due to the risks associated with immunosuppressive agents such as Ciclosporine, or Cyclophosphamide we decided to initiate a therapy with 50 mg/day Elthrombopaq, a TPO receptor agonist available for treatment of immune thrombocytopenia (ITP), which was successfully used in a patient with AATP and systemic lupus erythematosus (SLE) (Cela et al., 2010). After several days of treatment with Elthrombopaq we initially observed a rise in thrombocyte count and a decreased tendency to bleed. Thrombocyte counts, however, fell again after several weeks and the dose subsequently increased by 25 to 75 mg/day. Laboratory follow-ups showed no improvement of the thrombocyte count after several weeks, necessitating another hospitalization and change of treatment to a combination therapy of Romiplostim and Ciclosporine, which again did not result in thrombocyte counts > 50/nl (Figure 2). Repeated bone marrow histology then showed fatty degeneration, absent megakaryopoiesis and little granulo/erythropoiesis. In accordance to that, peripheral blood analysis revealed progressive leukopenia, as well as anemia, necessitating further erythrocyte transfusions. Finally, the diagnosis of aplastic anemia was made. Further workup showed inconspicuous JAK2-exon12 sequencing and cytomegalovirus infection was ruled out. Sequencing of ASXL1, CBL, DNMT3A, EZH2, RUNX, SF3B1, SRSF2, TET2, TP53, U2AF1, ZRSR2 did not reveal any mutations other than SNPs in TP53 and EZH2. BCOR, BCORL1, DNMT3A sequencing was uneventful.

With regards to the progressive leukopenia, as well as aplastic anemia further therapy with an anti-CD20 antibody was deemed unsuitable at that time point and work-up for allogeneic stem cell transplantation has been started.

Discussion

The distribution of AATP among gender is equal, but differs according to age. While most affected females are in the 40–60 age, more men are affected at both ends of age distribution (Figure 1) with a peak in the 60ties.

Analysis of published reports reveals a success rate of <20% using either corticoids or IVIG alone. Treatment with Ciclosporine yielded a response rate of about 50% compared to 80% with anti-thymocyte globulin (ATG). Two cases were successfully treated with anti-CD20 antibodies, while there is only one case report of successful allogeneic stem cell transplantation in a patient refractory to multiple immunosuppressive agents and one report on successful use of Mycophenolate Mofetil (Bulchandani et al., 2007) (Table 2). In scarce reports, anti-CD20 antibodies and TPO receptor agonists showed 100% response (N=2 each).

With less than a 20% response rate, corticoids and IVIG are insufficient as first line therapy in AATP. Treatment with ATG seems promising, but carries the risk of allo-immunization and requires in-hospital stay. Ciclosporine on the other
hand greatly increases the rate of infections, as well as the risk of liver damage necessitating close monitoring of liver enzymes and kidney function. Whether anti-CD20 therapy represents a better therapeutic option remains to be assessed. Due to the serious side effects accompanied with an allogeneic stem cell transplantation, we feel that this treatment option should be reserved for those in certain age, refractory to more than one therapeutic option other than corticoids or IVIG and absence of significant comorbidities. TPO receptor agonists seem to be a reasonable first line therapy due to their mechanism of action even despite our negative result.

Our case report shows the importance of complete diagnostics in patients presenting with apparent immune thrombocytopenia and their regular follow-ups. It unrolls the lack of an optimal treatment algorithm for AATP and depicts the importance of case reports for this entity in order to assess treatment success with different therapeutic regimens, since prospective randomized clinical studies are difficult to perform due to the rarity of the disease.

References

Table 2 – Percent response to specific therapy

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Response rate (%)</th>
<th>N</th>
</tr>
</thead>
<tbody>
<tr>
<td>Corticoids</td>
<td>9</td>
<td>22</td>
</tr>
<tr>
<td>Intravenous immunoglobulin</td>
<td>14</td>
<td>7</td>
</tr>
<tr>
<td>Ciclosporine</td>
<td>50</td>
<td>6</td>
</tr>
<tr>
<td>Anti-thymocyte globulin</td>
<td>80</td>
<td>5</td>
</tr>
</tbody>
</table>

Progression of AATP into Aplastic Anemia


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